

Review article

DNA methylation in phenylketonuria: a narrative review

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Received: January 2018; Accepted: April 2018

Abstract: Phenylketonuria (PKU) occurs due to a mutation in the gene encoding phenylalanine hydroxylase, which results in inability to convert phenylalanine into tyrosine. DNA methylation is an important epigenetic modification of the genome. Many human diseases have been detected to be related to aberrant DNA methylation. Investigating the leukocytes of PKU patients exposed to phenylalanine has shown a wide range of methylation, which indicates DNA methylation changes as a biochemical marker. In this article, we reviewed evidence of DNA methylation in pathophysiology of PKU.

Keyword: DNA methylation; Phenylketonuria; phenylalanine

Cite this article as: Siyahmarzkouhi F, Sarveazad A, Siyahmarzkouhi Z, Dindarlou R. DNA methylation in phenylketonuria: a narrative review. J Med Physiol. 2018; 3: e3.

1. Introduction

Phenylketonuria (PKU), known as the most common inborn error of amino acid metabolism, is due to a mutation in the gene encoding phenylalanine hydroxylase, which results in inability to convert phenylalanine into tyrosine, and consequently a high level of phenylalanine in the blood (1, 2). This disease is a recessive autosomal genetic disorder caused by a deficiency in phenylalanine hydroxylase that leads to intellectual disability if not treated.

Maternal PKU (MPKU) often affects the fetus and causes congenital heart defects and microcephaly. Management of MPKU has primarily concentrated on controlling the high level of phenylalanine in the blood (3). Recently, stem cell therapy (4) such as other disease (5-15) and antioxidant therapy (16-21) present for PKU management. The deficiency of phenylalanine hydroxylase activity in the liver is due to a mutation in chromosome 12, which causes metabolic disorders. The locus of the human chromosome 12q23.2 contains a gene that codes

for the phenylalanine hydroxylase enzyme, which has hundreds of alleles, often homozygous phenotypes (22). Investigation has shown that diet alone is not enough to cure this disease (23).

2. Epidemiology

PKU is transmitted as an autosomal recessive trait and has a collective prevalence of about one per 10,000 people, therefore, 2 percent of people carry PKU gene (24, 25). The incidence of PKU was reported to be 1 per 1500 birth in the USA (26). It varies around the world, but its average frequency is 1 in 10000 (27).

3. Etiology

This congenital disorder is caused by a deficiency in phenylalanine hydroxylase (PAH) (25), the enzyme which converts phenylalanine to tyrosine, consequently, the conversion of phenylalanine to tyrosine is interrupted (2, 25). PAH is a liver enzyme that increases the hydroxylation rate of 1-phenylalanine to 1-tyrosine by using tetrahydrobiopterin (BH4) cofactor (26). The polyhydroxyalkanoate (PHA) gene is about 90 kb and contains 13 exons encoding 451 amino acids for synthe-

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sis of the protein (28). PKU gene is located on chromosome 12q23.1, with more than 500 mutations reported in PAH gene, most of which are point mutations (29, 30). Loci that cover phenylalanine hydroxylase are 1.5 mbp. Exons in the PAH gene make approximately 2.88% of the genomic sequence lying between the start codon and 3' poly A. The longest exon and intron are exon 13 (892 bp) and intron 2 (17.87 bp), respectively. The shortest ones are exon 9 (57 bp) and intron 10 (556 bp). The PHA gene contains 40.7% GC in its sequence (31). The most common mutation with a relative frequency of 42% is the substitution of arginine with tryptophan (27). PKU occurs only due to defective activity of the PAH enzyme that is expressed only in the liver. The normal phenylalanine concentration is usually between 50 and 120 $\mu\text{mol/L}$ (32).

In case of PKU, genotype is the more important factor compared to phenylalanine intake. However, people may carry this genotype, but the disease will only be developed by phenylalanine intake and the severity of disease can be improved by diet control (33). Genetics and epigenetics are likely to play a significant role in onset of diseases (34). Aberrant methylation in CpG island of the promoter results in the silencing of argininosuccinate synthetase (ASS) and consequently, arginine biosynthesis disorders. The changes in expression of ASS enzyme affect vascular contraction and metabolic functions (35). ASS is the rate-limiting enzyme in arginine synthesis pathway. The cell is able to convert citrulline to arginine via this metabolic pathway.

Investigating the leukocytes of PKU patients exposed to

phenylalanine has shown a wide range of methylation, which indicates DNA methylation changes as a biochemical marker. Studies have shown that a gene that plays a role in development of the nervous system has a particular methylation pattern that affects the expression of downstream genes. GTGTG demethylation and GTGC / TG, PAH partial methylation have been reported in healthy people, indicating that they are not pathogenic alleles. The analysis of GPX3 promoter DNA methylation (GPX3) indicated increased production of primary radicals (36). Phosphodiester bond guanine positions overlap with CCAAT box/metal. The response element (CGATTGGCTG) of the active GPX3 promoter is analyzed by the oxidative stimulus. The CCAAT-box / metal interference is an interesting response element. Because this response element can not only be activated by reactive oxygen species (ROS) induced phenylalanine, it can also be activated by metal ions resulting from ROS hemostatic disorder by creating an imbalance (37, 38). Studies on GTGTG methylation and partial methylation of GTGC/TG PHA have shown that these alleles are not pathogenic (38).

It has been shown that ASS methylation can be detected before the obvious PKU symptoms (35). The relative frequency of mutations in PKU can be estimated as 0.01 in the population, but among these mutations, C1222 C>T allele (P.R408W) has the highest rate. Studies have shown that deamination-methylation of 5-methylcytosine (5mC) has an important role in CpG mutations in the human genome (39).

Table 1: Studies for methylation process in PKU

Author, year, country	Number of control and treated patients	Treatment protocols	Main findings
Murphy, et al., 2006, Canada	3 PKU patients / 12 normal controls	Antonarakis and the Nomenclature Working Group 1998, Invitrogen; Carlsbad, CA Wizard DNA Clean-up System, Promega; Madison, WI	Methylation-mediated deamination of c.1222mC can be a source of recurring c.1222C>T, p.R408W alleles on different background haplotypes.
Dobrowolski, et al., 2014, United States	19 fetuses of PHE unrestricted mice	Harlan Laboratories	In utero exposure to phenylalanine toxicity is associated with aberrant DNA methylation in the brain.
Dobrowolski, et al., 2015, United States	2 classical PKU patients / 5 control brain tissues	A protocol was obtained through the University of Pittsburgh	Abnormal DNA methylation is presented in PKU patients and is influenced by phenylalanine exposure.

PKU: Phenylketonuria

4. DNA methylation and PKU

Regarding methylation process in PKU, there are limited studies (Table 1). Murphy et al., have stated that concentrated deamination on methylation of 5mC is implicated in CpG mutation. In fact, methylated cytosine in mCpG nucleotide field may induce by itself and initiate C>T replacement. It has been shown that PHA allele is probably a repeated mutation C.1222 C>T, containing a methylated cytosine. There is the similar pattern of methylation in CpG dinucleotide. In homozygous patient samples of p.R408W mutation, T has been recorded in c.1222 nucleotide (A in complementary DNA). In general, based on these evidence, it has been hypothesized that methylation-mediated deamination of c.1222 mc is the main mechanism involved in the repeat of c.1222 C>T, P.r408w alleles in different background haplotypes. The highest observed amounts of c.1222 C>T, P.R408W is the main reason for outbreak of PKU disease in the Caucasian population (40).

Dobrowlski et al., stated that phenylalanine is known as a toxic component in PKU patients, but the exact mechanism of its toxicity is unclear. They concluded that the pattern of DNA methylation may be a critical biomarker relating to historic phenylalanine exposure. These data may improve quality of therapy (41).

Findings of Scriver et al., showed that altered DNA methylation in brain due to phenylalanine toxicity is fetal. In the mentioned research, PKU mice were used as a model. The diet with limited phenylalanine led to the levels of PHE \leq 150 μ m in blood, while in unlimited condition, the concentration of phenylalanine was 100 μ m. Assessment of methylation process in promoters of 17 samples revealed that in contrast to MPKU, expression of these genes was reduced in PKU (31).

This opinion that "phenylalanine toxicity may modify DNA methylation in brain tissue" is a key hypothesis for clarifying pathophysiological mechanisms. Various kinds of individual cell types have different sensitivities to phenylalanine poisoning, resulting in different responses in methylome and consequently, changes in gene expression occur. The exact alterations in gene expression are not clear (42). It is obvious that more research is needed to clarify the exact implicated mechanisms.

5. Conclusion

Data on the role of DNA methylation in pathology of phenylketonuria is very limited. Studies have suggested that DNA methylation may play a role in the mechanism of phenylalanine toxicity in PKU. Accordingly, some studies have suggested DNA methylation as a possible biomarker relating to historic phenylalanine exposure.

6. Acknowledgment

None.

7. Conflict of interest

No conflict of interest was declared.

8. Funding source

None.

9. Author contribution

All authors passed four criteria for authorship contribution based on recommendations of the International Committee of Medical Journal Editor.

10. Reference

- Eiken H, Odland E, Boman H, Skjelkvåle L, Engebretsen L, Apold J. Application of natural and amplification created restriction sites for the diagnosis of PKU mutations. *Nucleic Acids Res.* 1991;19(7):1427-30.
- Woo SL, Lidsky AS, Güttler F, Chandra T, Robson KJ. Cloned human phenylalanine hydroxylase gene allows prenatal diagnosis and carrier detection of classical phenylketonuria. *Nature.* 1983;306(5939):151-5.
- Yano S, Moseley K, Bottiglieri T, Arning E, Azen C. Maternal Phenylketonuria International Collaborative Study revisited: evaluation of maternal nutritional risk factors besides phenylalanine for fetal congenital heart defects. *J Inherit Metab Dis.* 2014;37(1):39-42.
- Strisciuglio P, Concolino D. New Strategies for the Treatment of Phenylketonuria (PKU). *Metabolites.* 2014;4(4):1007-17.
- Akhkand SS, Amirzadeh N, Nikougoftar M, Alizadeh J, Zaker F, Sarveazad A, et al. Evaluation of umbilical cord blood CD34+ hematopoietic stem cells expansion with inhibition of TGF- β receptorII in co-culture with bone marrow mesenchymal stromal cells. *Tissue Cell.* 2016;48(4):305-11.
- Amini N, Vousooghi N, Hadjighassem M, Bakhtiyari M, Mousavi N, Safakheil H, et al. Efficacy of Human Adipose Tissue-Derived Stem Cells on Neonatal Bilirubin Encephalopathy in Rats. *Neurotox Res.* 2016;29(4):514-24.
- Babahajian A, Shamseddin J, Sarveazad A. Stem cell therapy in fecal incontinence: a narrative review. *J Med Physiol.* 2017;2(1):2-9.
- Faghihi F, Mirzaei E, Sarveazad A, Ai J, Barough SE, Lotfi A, et al. Differentiation potential of human bone marrow mesenchymal stem cells into motoneuron-like cells on electrospun gelatin membrane. *J Mol Neurosci.* 2015;55(4):845-53.
- Goudarzi F, Sarveazad A, Mahmoudi M, Mohammadalipour A, Chahardoli R, Malekshah OM, et al. Combined effect of retinoic acid and calcium on the in vitro differentiation of human adipose-derived stem cells to adipocytes. *Arch Physiol Biochem.* 2017:1-10.

10. Nakhjavan-Shahraki B, Yousefifard M, Oraili A, Sarveazad A, Hosseini M. Meta-analysis of neuron specific enolase in predicting pediatric brain injury outcomes. *EXCLI J*. 2017;16:995.
11. Sarvandi SS, Joghataei MT, Parivar K, Khosravi M, Sarveazad A, Sanadgol N. In vitro differentiation of rat mesenchymal stem cells to hepatocyte lineage. *Iran J Basic Med Sci*. 2015;18(1):89-97.
12. Sarveazad A, Babahajian A, Bakhtiari M, Soleimani M, Behnam B, Yari A, et al. The combined application of human adipose derived stem cells and Chondroitinase ABC in treatment of a spinal cord injury model. *Neuropeptides*. 2017;61:39-47.
13. Sarveazad A, Babahajian A, Yousefifard M. Human Adipose-Derived Stem/Stromal Cells from Children or Adults? *Int J Pediatr*. 2017;5(12):6779-80.
14. Sarveazad A, Bakhtiari M, Babahajian A, Janzade A, Fallah A, Moradi F, et al. Comparison of human adipose-derived stem cells and chondroitinase ABC transplantation on locomotor recovery in the contusion model of spinal cord injury in rats. *Iran J Basic Med Sci*. 2014;17(9):685-93.
15. Sarveazad A, Newstead GL, Mirzaei R, Joghataei MT, Bakhtiari M, Babahajian A, et al. A new method for treating fecal incontinence by implanting stem cells derived from human adipose tissue: preliminary findings of a randomized double-blind clinical trial. *Stem Cell Res Ther*. 2017;8(1):40.
16. Mazzola PN, Karikas GA, Schulpis KH, Dutra-Filho CS. Antioxidant treatment strategies for hyperphenylalaninemia. *Metab Brain Dis*. 2013;28(4):541-50.
17. Asadi MH, Zafari F, Sarveazad A, Abbasi M, Safa M, Koruji M, et al. Saffron improves epididymal sperm parameters in rats exposed to cadmium. *Nephrourol Mon*. 2014;6(1):e12125.
18. Babahajian A, Rasouli H, Katebi M, Sarveazad A, Soleimani M, Nobakht M. Effect of human chorionic gonadotropin and vitamin E on cellular density of CA1 hippocampal area, learning ability and memory, following ischemia-reperfusion injury in mice. *J Gorgan Uni Med Sci*. 2013;15(4):23-8.
19. Bahadoran H, Naghii M, Mofid M, Asadi M, Ahmadi K, Sarveazad A. Protective effects of boron and vitamin E on ethylene glycol-induced renal crystal calcium deposition in rat. *Endocr Regul*. 2016;50(4):194-206.
20. Sarveazad A, Babahajian A, Yari A, Goudarzi F, Soleimani M, Nourani M. Neuroprotective Role of Trolox in Hippocampus after Ischemia Reperfusion Injury in Mouse. *Int J Vitam Nutr Res*. 2017;1(1):1-7.
21. Yari A, Sarveazad A, Asadi E, Raouf Sarshoori J, Babahajian A, Amini N, et al. Efficacy of Crocus sativus L. on reduction of cadmium-induced toxicity on spermatogenesis in adult rats. *Andrologia*. 2016;48(10):1244-52.
22. Murphy B, Scriver C, Singh S. CpG methylation accounts for a recurrent mutation (c. 1222C>T) in the human PAH gene. *Hum Mutat*. 2006;27(9):975-.
23. Bollati V, Baccarelli A. Environmental epigenetics. *Heredity (Edinb)*. 2010;105(1):105-12.
24. Lidksy A, Robson K, Thirumalachary C, Barker P, Ruddle F, Woo S. The PKU locus in man is on chromosome 12. *Am J Hum Genet*. 1984;36(3):527-33.
25. Lidksy A, Güttler F, Woo SC. Prenatal diagnosis of classic phenylketonuria by DNA analysis. *Lancet*. 1985;325(8428):549-51.
26. Senemar S, Ganjekarimi H, Fathzadeh M, Tarami B, Bazrgar M. Epidemiological and clinical study of Phenylketonuria (PKU) disease in the National Screening Program of Neonates, Fars province, Southern Iran. *Iran J Public Health*. 2009;38(2):58-64.
27. O'Neill C, Eisensmith R, Croke D, Naughten E, Cahalane S, Woo S. Molecular analysis of PKU in Ireland. *Acta Paediatr*. 1994;83(s407):43-4.
28. DiLella AG, Marvi J, Brayton K, Woo SL. An amino acid substitution involved in phenylketonuria is in linkage disequilibrium with DNA haplotype 2. *Nature*. 1987;327(6120):333-6.
29. Vockley J, Andersson HC, Antshel KM, Braverman NE, Burton BK, Frazier DM, et al. Phenylalanine hydroxylase deficiency: diagnosis and management guideline. *Genet Med*. 2013;16(2):188-200.
30. Matalon R, Koch R, Michals-Matalon K, Moseley K, Surendran S, Tyring S, et al. Bipterin responsive phenylalanine hydroxylase deficiency. *Genet Med*. 2004;6(1):27-32.
31. Scriver CR. The PAH gene, phenylketonuria, and a paradigm shift. *Hum Mutat*. 2007;28(9):831-45.
32. Ramaswami U, Smith I. Phenylketonuria. *Current Paediatrics*. 1997;7(4):251-5.
33. Bird AP, Wolffe AP. Methylation-induced repression—belts, braces, and chromatin. *Cell*. 1999;99(5):451-4.
34. Wright RO. New morbidities: new challenges. *Curr Opin Pediatr*. 2009;21(2):220-1.
35. Wang D, Zhang H, Liang J, Li X, Feng X, Wang H, et al. Allogeneic mesenchymal stem cell transplantation in severe and refractory systemic lupus erythematosus: 4 years of experience. *Cell Transplant*. 2013;22(12):2267-77.
36. Anderson PJ, Leuzzi V. White matter pathology in phenylketonuria. *Mol Genet Metab*. 2010;99:S3-S9.
37. Walter JH. Vitamin B 12 deficiency and phenylketonuria. *Mol Genet Metab*. 2011;104:S52-S4.
38. Escueta S, Schanzer A, Farhadi S, Metz T, Zeyda M, Möslinger D, et al. Demethylation of the promoter region of GPX3 in a newborn with classical phenylketonuria. *Clin Biochem*. 2017;50(3):159-61.
39. Billings PR, Kohn MA, De Cuevas M, Beckwith J, Alper JS, Natowicz MR. Discrimination as a consequence of genetic testing. *Am J Hum Genet*. 1992;50(3):476.
40. Murphy BC, Scriver CR, Singh SM. CpG methylation accounts for a recurrent mutation (c.1222C>T) in the human PAH gene. *Human Mutation*. 2006;27(9):975-.
41. Dobrowolski SF, Lyons-Weiler J, Spridik K, Biery A, Breck J, Vockley J, et al. Altered DNA methylation in PAH deficient phenylketonuria. *Mol Genet Metab*. 2015;115(2-3):72-7.
42. Dobrowolski SF, Lyons-Weiler J, Biery A, Spridik K, Vockley G, Kranik E, et al. Methylome repatterning in a mouse model of Maternal PKU Syndrome. *Mol Genet Metab*. 2014;113(3):194-9.